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Investigation of the effects of Sugammadex and Rocuronium on rat lung tissue

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Copyright © 2024 The author(s) - Available online at www.ncmrjournal.com.tr. This is an open access article distributed under the terms of Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International License.***Address for correspondence:****Zümürüt DOGAN**Department of Anatomy, Adiyaman University,
Faculty of Medicine, Adiyaman, Türkiye**Mail:** byozumrut@yahoo.com**Orcid:** 0000-0001-7131-2317**DOI:** 10.5281/zenodo.11652915**Abstract**

In this study, sugammadex of used to reverse rocuronium used for surgical operations aimed to be examined effects on the lungs that expel anesthetic agents from the body. The possible mechanism of oxidative damage and the effects of sugammadex on lung morphology are still not understood. Four groups were formed from 32 adult Sprague-Dawley male rats as pure control, control group, sugammadex, and sugammadex-rocuronium. After drug administration, lung tissues were assessed histopathologically and biochemically for oxidative damage. A statistically significant difference was observed between the groups regarding glutathione (GSH) levels in the rat lung. It was observed that rocuronium increased the malondialdehyde (MDA) value, which is considered an indicator of free radical damage in tissues. The

sugammadex-rocuronium complex can cause oxidative stress in lung tissue. In group sugammadex, dense fibrosis and lymphoid tissue were found in the connective tissue, while group sugammadex-rocuronium had little fibrosis and lymphoid tissue. In group sugammadex-rocuronium, thickening of the alveolar wall was detected. The effect of sugammadex application on lung tissues has been demonstrated by biochemical and histopathologic data. Rocuronium caused an increase in mast cells in lung tissue. Sugammadex suppressed this increase caused by rocuronium and caused a decrease in mast cells in the lung tissue. Although these data were obtained as a result of experimental studies, we think that they will make a significant contribution to the anesthesia and reanimation patient treatment protocols in the clinic. However, new studies are necessary to determine the toxic effects of sugammadex and sugammadex-rocuronium complex.

Keywords: Sugammadex; rocuronium; lung morphology; oxidative stres; rat

Introduction

Neuromuscular blockade (NMB) is routinely a serious part of general anesthesia [1]. Rocuronium is available for use as a neuromuscular blocking agent that is indicated for routine or rapid intubation in patients under general anesthesia. Non-depolarizing Rocuronium is widely used as a neuromuscular blocker in general anesthesia surgeries. Sugammadex is frequently used as a rocuronium reversal agent [2]. The development of sugammadex and its introduction into clinical practice is a significant advance in the treatment of NMB. Sugammadex is a binder developed specifically as a reversal agent for the aminosteroid NMBs and rocuronium [3, 4]. Sugammadex was developed as a reversal without the limitations of anticholinesterase drugs. It first became known in the scientific community in 2002. Since then, research on this drug has gained momentum through animal and human experiments [5, 6]. It has taken its place among drugs as the first selective relaxant-binding agent based on cyclodextrin. It is known that it has been approved for clinical use in seventy countries around the world in the past years [7]. The sugammadex molecule in the Sugammadex-rocuronium complex is known to repair quickly and safely deep neuromuscular blockade caused by rocuronium binding to plasma [8,]. It is a modified γ -cyclodextrin developed from cyclodextrins used to dissolve steroids. It is a drug that selectively binds to steroidal neuromuscular blockers. This binding encapsulates and inactivates NMBs. The new compound formed is inactive and is removed from the body in accordance with the pharmacokinetic properties of sugammadex [5, 9]. The lungs are an organ that plays a role in the elimination of anesthetic agents from the body [10, 11]. The majority of relevant clinical studies have remained focused on evaluating its effectiveness against NMB [12], because of Sugammadex is specifically produced to reverse the blockade caused by rocuronium [3, 4]. Sugammadex's clinical use has increased due to its rapid and safe recovery. Although many studies emphasize that sugammadex is effective and safe, some studies have revealed potential risks [12]. Since Sugammadex is a relatively new drug [13], its histomorphological effects and oxidative stress mechanism in lung tissue have not yet been revealed. In this study, the possible effects of sugammadex and sugammadex-rocuronium complex on the possible oxidative damage mechanism and lung morphology were investigated.

Materials And Methods

Animals and Study Characteristics

The study was started after approval was received by the

Adiyaman University Animal Experiments Local Ethics Committee (Approval number: ADYU-HADYEK: 2019_042). Experiments were performed in accordance with the National Institutes of Health Guidelines for the Care and Use of Laboratory Animals, and required sensitivity was taken into account.

Rats kept at room temperature of 22-25 °C were fed daily with a standard pellet diet and water ad libitum. The humidity of the room was set to 50-55%. They were subjected to a 12-hour (06.00-18.00) light-dark cycle using cool white fluorescent lamps in order not to disrupt the natural light-dark cycle. 32 adult Sprague-Dawley male rats weighing between 300-350 g were divided into four equal groups by random selection. The groups consisted of the purine group, the control group, the sugammadex group, and the sugammadex-rocuronium group. The purine group was created from animals that did not undergo surgical treatment to obtain baseline values. Rats in our control group were injected intravenously (IV) with 16 mg/kg 0.9% isotonic saline. The Sugammadex group was created by giving 16 mg/kg IV Sugammadex (Bridion®; Schering-Plough Corporation, Oss, Netherlands) to the rats. The group that received 16 mg/kg IV sugammadex injection and three minutes later 1 mg/kg IV rocuronium (Esmeron®; Organon, Istanbul, Turkey) injection was determined as the sugammadex-rocuronium group. All injection procedures were performed through the tail vein. Drug dosage amounts were made according to Bostan et al. [14] and Pühringer et al. studies of [15] references. Three days after the experimental procedures, lung tissue was removed under ketamine/xylazine anesthesia.

Biochemical Procedure

The dissected lung tissue was first washed with physiological saline at +4°C. According to the cold chain principle, the washed tissue was stored in Eppendorf tubes at -70°C until analysis. For MDA and GSH measurements, tissue homogenates were first obtained from tissue samples. Tissue homogenates were created with the help of a cold 0.15 M KCl (10%, w/v) homogenizer. The Uchiyama method was used as a reference for MDA analysis, and studies were carried out using this method [16]. For GSH analysis, Ellman's [17] method was referenced, and applications were made accordingly.

The MDA test, which stands out with the increase in the amount of free oxygen radicals, is an indicator of the presence of oxidative stress. The pink-colored product is made by reading the pink-colored product from the N-butanol phase at 535 and 520 nm due to the reaction with thiobarbituric at 95°C.

Table 1. Lung biochemical parameters in all groups (n=8)

Parameter	Purine Control	Control	Sugammadex	Sugammadex+Rocuronium
GSH (nmol/g)	820.75±84.74	767.15±75.75	960.19±137.94 ^{a,b,c}	724.85±84.72
MDA (nmol/g)	1983.89±281.91	1962.26±246.38	1620.12±271.99 ^d	2012.74±829.88

^aP<0.05 compared to Purine Control P:0,041.
^bP<0.05 compared to Control P:0,003.
^cP<0.05 compared to Sugammadex+Rocuronium P :0,001.
^dP<0.05 compared to Sugammadex+Rocuronium P :0,004.

To create the homogen, lung tissue was homogenized with 10% trichloroacetic acid. The resulting homogenate was centrifuged, and an equal volume of 0.67% thiobutyric acid was added to the resulting supernatant. Incubation was carried out with 90°C boiling water for 15 minutes. Centrifugation was applied to the cooled supernatant. MDA concentrations were studied in lung tissue under 532 nm absorbance, and calculations were made using nmol/g tissue. GSH, which reduces harmful peroxides such as lipids and hydrogen peroxide, is an antioxidant enzyme. This enzyme converts reduced glutathione into oxidized glutathione, which is a marker of oxidative stress. 5,5'-dithiobis-2-nitrobenzoic acid is added into the analysis tubes containing the lung tissue. A reaction occurs between this acid and the sulfhydryl groups contained in glutathione. From this reaction, a yellow-greenish product content occurs. GSH concentration density is determined by measuring the absorbance of this product at 410 nm wavelength with a spectrophotometer. The study data are given in nmol/g.

Histopathological Procedure

The lung tissues of the experimental groups were placed in a 10% formaldehyde solution so that the groups would not be mixed. Fixation was applied for one week. Tissues were cleared of fixation solution by washing under tap water. Paraffin blocks were created adhering to the principle of routine histological tissue monitoring during the paraffin block preparation phase. The paraffin blocks we prepared were divided into sections with a thickness of 7 microns. The obtained paraffin sections were stained with Masson trichrome and toluidine blue dyes. Images obtained through a microscope with a digital camera attachment (Carl Zeiss brand Axiocam ERc5 model) were evaluated histomorphologically.

Statistical Tests

The statistical study was carried out using the Statistical Package for the Social Sciences 22.0 program (SPSS Inc., Chicago, IL, USA). First of all, the normal distribution of the data was evaluated with the single sample Kolmogorov-Smirnov test. A one-way ANOVA test was used because our groups were independent of each other, and the data showed normal distribution. All statistical analyses were considered with a 95% confidence interval and are expressed as mean ± standard deviation (SD). The significance value was taken as p<0.05.

Results

Biochemical Evaluation

As a result of statistical measurements of GSH levels in rat lungs, purine 820.75±84.74 nmol/g in the control group, 767.15±75.75 nmol/g in the control group, 960.19±137.94 nmol/g in the sugammadex group, in the sugammadex-rocuronium group 724.85±84.72 nmol/g found as wet tissue. MDA, symbol of free radical damage and lipid peroxidation, in the purine control group, 1983.89±281.91 nmol/g, 1962.26±246.38 nmol/g in the control group, 1620.12±271.99 nmol/g in the sugammadex group and 2012.74±829.88 nmol/g in the sugammadex-rocuronium group found as wet tissue (Table 1).

Histopathological Evaluation

Lung tissue was found to have a normal histomorphological appearance in the images of groups 1 and 2 in the examinations made. No pathological findings were found in the epithelium, lamina propria, muscle, and adventitia layer of the bronchial wall. Alveolar walls and vascular structures were normal (figure 1, 1 a-c, 2 a-c). There was a low

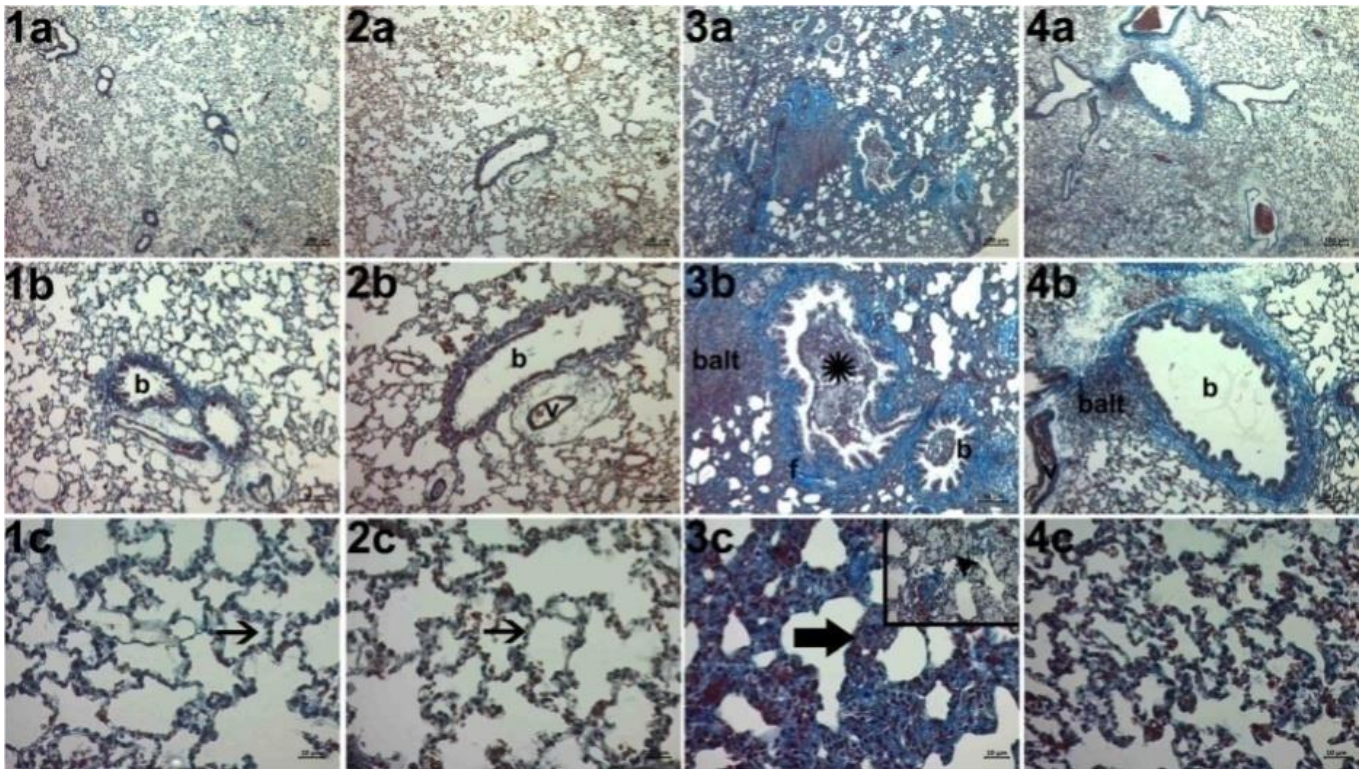


Figure 1. 1a, 1b and 1c; group 1 images of x4-x10-x40 magnifications, respectively; 2a, 2b and 2c; images of x4-x10-x40 magnifications, respectively; 3a, 3b and 3c; group 3 images at x4-x10-x40 magnifications; 4a, 4b and 4c; group 4 images of x4-x10-x40 magnifications (Staining masontrichrome); b; bronchiole, v; blood vessel, f; fibrosis, ax; lymphoid tissue, star; exudate, thin black arrow; normal alveolar wall, thick black arrow; collapse in the alveolar wall, black arrowhead; macrophage cell.

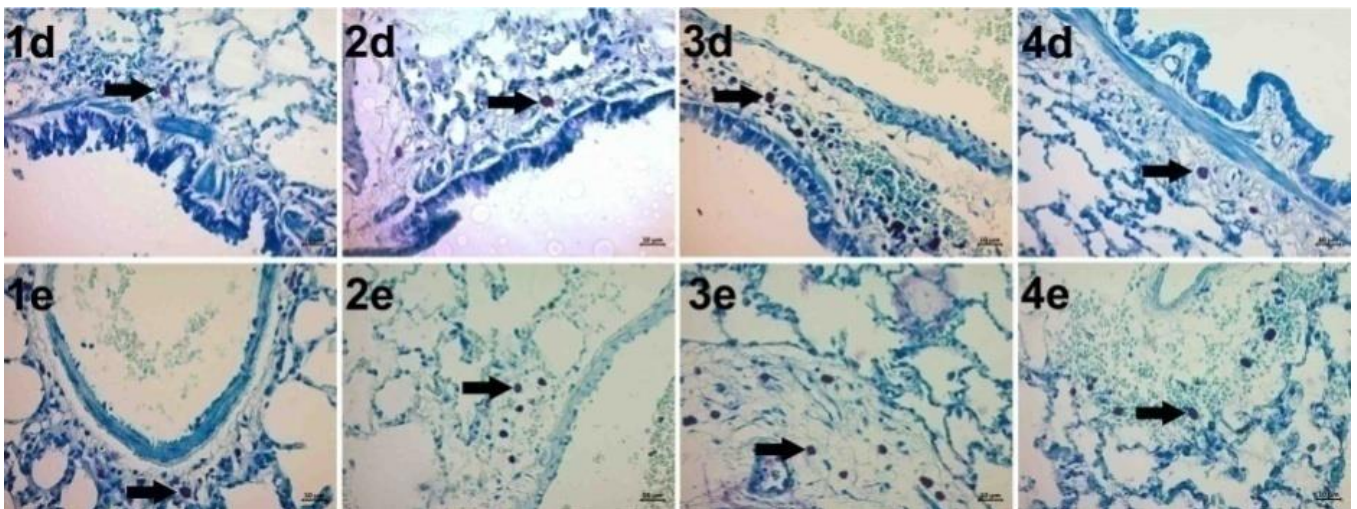


Figure 2. 1d and 1e; images of group 1 at x40 magnification; 2d and 2e; images of group 2 at x40 magnification; 3d and 3e; images of group 3 at x40 magnification; 4d and 4e; images of group 4 at x40 magnification; (Staining toluidine blue); black arrow; mast cells

density of mast cells around the bronchiole and vessel (figure 2, 1 d,e - 2 d,e). In the examinations of group 3, the epithelium of the bronchiole walls was normal, but there was fibrosis and dense lymphoid tissue in the connective tissue. In addition, findings of exudate were found in the lumen of the bronchiole. There was a collapse in the alveolar wall, dilatation of the alveolar capillaries, and dense macrophage cells (figure 1, 3 a-c). In addition, dense mast cells were seen around the bronchiole and vessel (figure 2, 3 d-e). In the examinations performed in Group 4, the epithelium of the bronchiole walls was normal, and there was little fibrosis and lymphoid tissue in the connective tissue. Thickening of the alveolar wall was detected (figure 1, 4 a-c). In addition, a decrease in the density of mast cells was observed around the bronchiole and vessel (figure 2, 4 d-e).

Discussion

Neuromuscular blocking agents (NMBAs) are important elements of general anesthesia to improve surgical outcomes [18]. There used to be only a reversal agent to revert NMB. This reversal agent had several disadvantages [2, 19]. Sugammadex is a new pharmaceutical agent [13]. It antagonizes steroidal neuromuscular blocking agents such as rocuronium [2, 19]. Sugammadex has also been reported to be safe for reversing NMB [20]. It came into use in December 2015 after receiving clinical use approval from the United States Food and Drug Administration (FDA). However, in preclinical studies, there is no clear information about whether sugammadex application has minor side effects [21]. Additionally, since it is a new drug, more studies are needed in this field [13].

When the case report of sugammadex-resistant rocuronium-induced respiratory paralysis in Marie-Tooth disease is examined, It has been reported that systematic studies are needed to clarify the conditions that place patients at risk of long-term stroke after receiving rocuronium in combination with sugammadex [22]. Therefore, we planned this study considering it was important with this experimental study we have conducted; important information is provided in the literature. Although these data were obtained as a result of experimental studies, we believe that they will make a significant contribution to anesthesia and reanimation patient treatment protocols in the clinic.

Alagöz et al. showed no significant histopathological changes become evident in the group in which they administered sugammadex at a dose of 4 mg/kg; however, when administered at a dose of 16 mg/kg in the group, widespread irregularity of muscle fibers, degeneration, and openings between muscle fibers stated that they found it [23].

We determined the dose of sugammadex in our experimental group as 16 mg/kg based on the information presented to us by Alagöz and his friends in their studies. In addition, the study of Bostan et al. emphasized that rats receiving only rocuronium showed more significant histopathological changes compared to rats receiving only sugammadex. Their explanation is that sugammadex not only has a minimal effect when applied alone but may also worsen rocuronium-induced histopathological degeneration.

The lung is one of the vital organs that ensures the continuity of the vital cycle. If it cannot perform its duty, it is impossible to continue life [24]. General anesthesia is known to have profound effects on lung function. However, it is difficult to generalize the effects of anesthesia on respiratory function [25]. The effect that NBAs can have on the lung tissue, which is such an important organ for vital functions, is very important. An organ that is so important for vital functions how it will affect is very important by NBAs used for anesthetic needs.

Anesthetic agents used during surgical operations can be determinants of oxidative stress. It has been reported that the resulting oxidative stress causes lung damage, is a trigger for many events in the organism and is also associated with various complications in terms of clinical effects [26]. It is known that an increase in oxidative stress causes both damage at the molecular level and disruption of the oxidant-antioxidant balance in the organism [27]. Pathological conditions caused by oxidative stress can cause temporary or permanent damage to tissues [28]. While a high MDA level is defined as one of the markers of oxidative stress [29], the increase in GSH level also indicates the antioxidant defense system against oxidative stress [30]. Therefore, we first studied tissue markers of oxidative stress mainly to test the protective role of Sugammadex. Our study revealed that rocuronium causes lung damage by increasing oxidative stress in the tissue, and Sugammadex plays a protective role by increasing the antioxidant system. Koc et al. reported that Sugammadex inhibits the increase in MDA levels and significantly suppresses the decrease in GSH levels. They said that Sugammadex might be useful in preventing oxidative stress [31].

Mast cells, which play a significant role in the occurrence of pathological processes, are cells located throughout the body, including the respiratory system organs and other organs [32, 33, 34]. It is also known that the lungs of a healthy adult individual contain many mast cells under normal conditions [35, 36]. Researchers have reported that both anaphylaxis due to Sugammadex and allergic reactions to the Sugammadex-

rocuronium complex may occur [37].

Yeşiltaş and his colleagues [38] stated that rocuronium and sugammadex were investigated in experimental studies on the liver and pancreas, but their effects on the lungs were neglected. They also demonstrated that rocuronium-induced allergic effects in rat lungs were alleviated by sugammadex. When they histomorphologically examined the rat lungs of the experimental group in which sugammadex was applied, they stated that there was a slightly thickened alveolar wall in some regions. They explained that the number of mast cells in rat lung tissue was higher in the experimental group to which they applied rocuronium compared to all experimental groups. In our study, we observed that Sugammadex decreased the number of mast cells, and rocuronium increased, as in the study conducted by Yeşiltaş et al. We observed a significant increase in the alveolar wall in the morphological examination of lung tissue after rocuronium application. Increased MDA and decreased GSH levels support the morphological data. Although we see that our study data supports the literature data, we think that more studies are needed. As stated by Yeşiltaş et al., we observed that the research on the histomorphological and biochemical effects of sugammadex and rocuronium complex on the lung in the literature is still limited and insufficient.

In 2021, Yeşiltaş et al. [38] reported that the effects of rocuronium and sugammadex on the lungs were neglected, and there were not enough studies. As a result of our literature searches, no study has been found on the lungs in this field since 2021. We see that work restrictions still continue. We underline that more studies are needed in this field, as emphasized by Yeşiltaş et al [38].

Conclusion

Sugammadex and sugammadex-rocuronium complex were determined by histomorphological and biochemical effects on rat lung tissue after NMB. It was concluded that the sugammadex application did not have a negative effect on lung tissues. Additionally, sugammadex can suppress rocuronium-induced anaphylaxis and allergy by reducing the number of mast cells in the lung tissue. It was found that rocuronium did not cause pathological consequences in lung tissue morphology but caused oxidative damage through GSH and MDA. Although our study results are first, new studies will be more effective in revealing the possible effects of sugammadex and sugammadex-rocuronium complex on lung tissue. Finally, we can say that this study, like all research, is limited. These limitations are due to the lack of similar studies in the literature. Including new studies in the literature and

increasing the number of studies will benefit future studies. Another limitation of our study results is that we need clinical data. It is impossible to say whether it correlates with clinical findings based on experimental study data. However, it is possible to say that it is a preliminary study for future research and contains important data.

Ethical Declarations

Adıyaman University Animal Experiments Local Ethics Committee (Approval number: ADYU-HADYEK: 2019_042).

Conflict of Interest Statement

The authors disclosed no conflict of interest during the preparation or publication of this manuscript.

Financial Disclosure

No financial support was received for the study.

Data availability statement

The authors confirm that the data supporting the findings of this study are available in the article.

Author Contributions

All of the authors declare that they have all participated in the design, execution and analysis of the paper and that they have approved the final version.

References

1. Lee S, Jang EA, Chung S, Kang DH, Park SM, Hong M, Kim J, Jeong S. Comparisons of surgical conditions of deep and moderate neuromuscular blockade through multiple assessments and the quality of postoperative recovery in upper abdominal laparoscopic surgery. *J Clin Anesth.* 2021;73:110338. doi: 10.1016/j.jclinane.2021.110338
2. Renew, J. R., et al. Clinical use of neuromuscular blocking agents in anesthesia. *UpToDate.* 2019.
3. Booi LH. Cyclodextrins and the emergence of sugammadex. *Anaesthesia.* 2009;64:Suppl:1:31-7. doi: 10.1111/j.1365-2044.2008.05868.x
4. Booi LH, van Egmond J, Driessen JJ, de Boer HD. In vivo animal studies with sugammadex. *Anaesthesia.* 2009;64:Suppl 1:38-44. doi: 10.1111/j.1365-2044.2008.05869.x
5. Yang LP, Keam SJ. Sugammadex: a review of its use in anaesthetic practice. *Drugs.* 2009;69(7):919-42. doi: 10.2165/00003495-200969070-00008
6. Peeters PA, van den Heuvel MW, van Heumen E, Passier PC, Smeets JM, van Iersel T, Zwiers A. Safety, tolerability and pharmacokinetics of sugammadex using single high doses (up to 96 mg/kg) in healthy adult subjects: a randomized, double-blind, crossover, placebo-controlled, single-centre study. *Clin Drug Investig.* 2010;30(12):867-74. doi: 10.1007/BF03256915

7. Andersson CK, Mori M, Bjermer L, Löfdahl CG, Erjefält JS. Novel site-specific mast cell subpopulations in the human lung. *Thorax*. 2009;64(4):297-305. doi: 10.1136/thx.2008.101683
8. Sorgenfrei IF, Norrild K, Larsen PB, Stensballe J, Ostergaard D, Prins ME, Viby-Mogensen J. Reversal of rocuronium-induced neuromuscular block by the selective relaxant binding agent sugammadex: a dose-finding and safety study. *Anesthesiology*. 2006;104(4):667-74. doi: 10.1097/0000542-200604000-00009
9. Yağan Ö, Taş N, Mutlu T, Hancı V. Comparison of the effects of sugammadex and neostigmine on postoperative nausea and vomiting. *Braz J Anesthesiol*. 2017;67(2):147-152. doi: 10.1016/j.bjane.2015.08.003
10. Behrooz A. Pharmacogenetics and anaesthetic drugs: Implications for perioperative practice. *Ann Med Surg (Lond)*. 2015;10(4):470-4. doi: 10.1016/j.amsu.2015.11.001
11. Nair AS. Pharmacogenomics of inhalational anesthetic agents. *Med Gas Res*. 2019;9(1):52-53. doi: 10.4103/2045-9912.254641
12. Lee W. The potential risks of sugammadex. *Anesthesia and Pain Medicine*. 2019;14(2): 117-122. doi: 10.17085/apm.2019.14.2.117
13. Muedra V, Rodilla V, Llansola M, Agustí A, Pla C, Canto A, Hernández-Rabaza V. Potential Neuroprotective Role of Sugammadex: A Clinical Study on Cognitive Function Assessment in an Enhanced Recovery After Cardiac Surgery Approach and an Experimental Study. *Front Cell Neurosci*. 2022;16:789796. doi: 10.3389/fncel.2022.789796
14. Bostan H, Kalkan Y, Tomak Y, Tumkaya L, Altuner D, Yılmaz A, Erdivanlı B, Bedir R. Reversal of rocuronium-induced neuromuscular block with sugammadex and resulting histopathological effects in rat kidneys. *Ren Fail*. 2011;33(10):1019-24. doi: 10.3109/0886022X.2011.618972
15. Pühringer FK, Rex C, Sielenkämper AW, Claudius C, Larsen PB, Prins ME, Eikermann M, Khuenl-Brady KS. Reversal of profound, high-dose rocuronium-induced neuromuscular blockade by sugammadex at two different time points: an international, multicenter, randomized, dose-finding, safety assessor-blinded, phase II trial. *Anesthesiology*. 2008;109(2):188-97. doi: 10.1097/ALN.0b013e31817f5bc7
16. Mihara M, Uchiyama M. Determination of malonaldehyde precursor in tissues by thiobarbituric acid test. *Anal Biochem*. 1978;86(1):271-8. doi: 10.1016/0003-2697(78)90342-1
17. Ellman GL. Tissue sulfhydryl groups. *Arch Biochem Biophys*. 1959;82(1):70-7. doi: 10.1016/0003-9861(59)90090-6
18. Fuchs-Buder T, Romero CS, Lewald H, Lamperti M, Afshari A, et al. Peri-operative management of neuromuscular blockade: A guideline from the European Society of Anaesthesiology and Intensive Care. *Eur J Anaesthesiol*. 2023;40(2):82-94. doi: 10.1097/EJA.0000000000001769
19. Herring WJ, Woo T, Assaid CA, Lupinacci RJ, Lemmens HJ, Blobner M, Khuenl-Brady KS. Sugammadex efficacy for reversal of rocuronium- and vecuronium-induced neuromuscular blockade: A pooled analysis of 26 studies. *J Clin Anesth*. 2017;41:84-91. doi: 10.1016/j.jclinane.2017.06.006
20. Carron M, Tessari I, Linassi F. Sugammadex compared with neostigmine in reducing postoperative pulmonary complications in older patients: a meta-analysis. *Br J Anaesth*. 2022;128(4):e259-e262. doi: 10.1016/j.bja.2021.12.038
21. Tobias JD. Current evidence for the use of sugammadex in children. *Paediatr Anaesth*. 2017;27(2):118-125. doi: 10.1111/pan.13050
22. Hiramatsu S, Moriwaki K, Nakao M, Tsutsumi YM. Rocuronium-induced respiratory paralysis refractory to sugammadex in Charcot-Marie-Tooth disease. *Can J Anaesth*. 2022;69(3):364-368. doi: 10.1007/s12630-021-02168-y
23. Alagöz A, Küçükgüçlü S, Boztaş N, Hancı V, Yuluğ E, Şişman AR. Effects of sugammadex on ischemia reperfusion in a rat extremity model. *Ulus Travma Acil Cerrahi Derg*. 2020;26(4):509-516. doi: 10.14744/tjtes.2019.12524
24. Erçin BS, Kılıç KD, Tiftikçiöğlü YÖ, Biçer A, Uyanıkgil Y, Thione A, Gürler T. Solid organ nakli ve vaskülarize kompozit allotransplantasyon: Dünü ve bugünü. *İstanbul Bilim Üniversitesi Florence Nightingale Transplantasyon Dergisi*. 2017;2(1):7-13. doi: 10.5606/fng.transplantasyon.2017.002
25. Levitzky MG. Effects of Anesthesia on Pulmonary Function. In: Levitzky M, McDonough K, Kaye A, Hall S. eds. *Clinical Physiology in Anesthetic Practice*. McGraw Hill; 2021. <https://accessanesthesiology.mhmedical.com/content.aspx?bookid=2979§ionid=249590743>
26. Bermede, O. Tek Akciğer Ventilasyonu Uygulanan Hastalarda Sevofluran ve Propofolün Pulmoner Oksidatif Stres Üzerine Etkisi. *Journal of Ankara University Faculty of Medicine*. 2020;73(2).
27. Sies H. Oxidative Stress: Concept and Some Practical Aspects. *Antioxidants (Basel)*. 2020;9(9):852. doi: 10.3390/antiox9090852
28. Zhang C, Wang N, Xu Y, Tan HY, Li S, Feng Y. Molecular Mechanisms Involved in Oxidative Stress-Associated Liver Injury Induced by Chinese Herbal Medicine: An Experimental Evidence-Based Literature Review and Network Pharmacology Study. *Int J Mol Sci*. 2018;19(9):2745. doi: 10.3390/ijms19092745
29. Kehrer JP, Biswal SS. The molecular effects of acrolein. *Toxicol Sci*. 2000;57(1):6-15. doi: 10.1093/toxsci/57.1.6
30. Jiménez-Fernández S, Gurpegui M, Garrote-Rojas D, Gutiérrez-Rojas L, Carretero MD, Correll CU. Oxidative stress parameters and antioxidants in patients with bipolar disorder: Results from a meta-analysis comparing patients, including stratification by polarity and euthymic status, with healthy controls. *Bipolar Disord*. 2021;23(2):117-129. doi: 10.1111/bdi.12980
31. Koç A, Kuyrukçuyıldız U, Gazi M, Caner Sayar A, Altuner D, et al. The effects of sugammadex on gastric ischemia-reperfusion injury in rats: Biochemical and histopathological evaluation. *Gen Physiol Biophys*. 2023;42(1):67-75. doi: 10.4149/gpb_2022049
32. Atiakshin D, Buchwalow I, SamoiloVA V, Tiemann M. Trypsinase as a polyfunctional component of mast cells. *Histochem Cell Biol*. 2018;149(5):461-477. doi: 10.1007/s00418-018-1659-8
33. Atiakshin D, Buchwalow I, Tiemann M. Mast cell chymase: morphofunctional characteristics. *Histochem Cell Biol*. 2019;152(4):253-269. doi: 10.1007/s00418-019-01803-6
34. Theoharides TC. Potential association of mast cells with coronavirus disease 2019. *Ann Allergy Asthma Immunol*. 2021;126(3):217-218. doi: 10.1016/j.anai.2020.11.003
35. Andersson CK, Mori M, Bjermer L, Löfdahl CG, Erjefält JS. Novel site-specific mast cell subpopulations in the human lung. *Thorax*. 2009;64(4):297-305. doi: 10.1136/thx.2008.101683
36. Erjefält JS. Mast cells in human airways: the culprit? *Eur Respir Rev*. 2014;23(133):299-307. doi: 10.1183/09059180.00005014
37. Asakura C, Iwasaki H. The use of succinylcholine after sugammadex reversal. *J Anesth*. 2016;30(5):915. doi: 10.1007/s00540-016-2203-4
38. Yeşiltaş S, Orhon ZN, Cakır H, Doğru M, Çelik MG. Does Sugammadex Suppress Allergic Inflammation Due to Rocuronium in Animal Model of Rat? *Allergol Immunopathol (Madr)*. 2021;49(3):91-99. doi: 10.15586/aei.v49i3.84